

LOCAL OPERATING PROCEDURE

# **NEONATAL SERVICES DIVISION**

Approved by Quality & Patient Safety Committee 17 September 2020

# HEEL PRICK FOR BLOOD SAMPLING

This Local Operating Procedure is developed to guide safe clinical practice in Newborn Care Centre (NCC) at The Royal Hospital for Women. Individual patient circumstances may mean that practice diverges from this Local Operating Procedure.

Using this document outside the Royal Hospital for Women or its reproduction in whole or part, is subject to acknowledgement that it is the property of NCC and is valid and applicable for use at the time of publication. NCC is not responsible for consequences that may develop from the use of this document outside NCC.

#### 1. AIM

• To collect capillary blood by correct heel stick technique

#### 2. PATIENT

Newborns

#### 3. STAFF

• Medical and nursing staff

#### 4. EQUIPMENT

- Gloves
- Gauze
- Appropriate Puncture Device
- 0.5% Chlorhexidine Swab
- Required Blood/Capillary Tubes
- 25% Oral Sucrose or breast milk

Device	Device Name	Device Type	Width x Depth (mm)	Intended Use <sup>4</sup>	
	BD Microtainer® Contact-Activated Lancet (Purple)	Puncture (needle)	30 G x 1.5 mm	Fingerstick – Low Flow (single drop) Demonstrates significantly less pain for your patients than comparable products*	Preterm (BSL only)
	BD Microtainer® Contact-Activated Lancet (Pink)	Puncture (needle)	21 G x 1.8 mm	Fingerstick – Medium Flow	Not used in NCC
	BD Microtainer® Contact-Activated Lancet (Blue)	Puncture (blade)	1.5 mm x 2.0 mm	Fingerstick – High Flow (500 µL from single puncture)	Not used in NCC
$\bigcirc$	BD Microtainer® Quikheel™ Lancet (Pink)	Incision (blade)	1.75 mm x 0.85 mm	Heelstick – Low Flow (premature infants) Low birth-weight babies or full-term infants where lower blood volume is required	Preterm (all other blood tests)
	BD Microtainer® Quikheel <sup>™</sup> Lancet (Teal)	Incision (blade)	2.5 mm x 1.0 mm	Heelstick – High Flow (infants) Full-term infants where higher blood volume is required	Term (all other blood tests)

Picture 1

## 5. CLINICAL PRACTICE

### Procedure:

- 1. Perform hand hygiene and clean blue tray.
- 2. Collect equipment. Select the appropriate puncture device (Picture 1).
- 3. Administer 25% sucrose or breast milk and provide comfort measures (eg. swaddling, skin contact)
- 4. Perform hand hygiene, apply gloves and prepare equipment.
- 5. Nominate an area for puncture on the foot on the medial or lateral plantar surface (Picture 2).
- 6. Select the surface area to puncture. Continue in a "stepping" ladder pattern from the first puncture for subsequent blood sampling.
- 7. Clean foot with 0.5% chlorhexidine wipe and allow to dry for 30 seconds.



2. LOCAL OPERATING PROCEDURE

**NEONATAL SERVICES DIVISION** 

Approved by Quality & Patient Safety Committee 17 September 2020

# HEEL PRICK FOR BLOOD SAMPLING cont'd

8. Puncture heel holding the puncture device at a 90 degree angle (Picture 3 & 4).



Picture 2

Picture 3

Picture 4

- 9. Wipe away first drop of blood with gauze.
- 10. Collect blood in correct order of draw (Picture 5), gently agitating tubes between each drop while avoiding scraping and scooping.

Or BD M	der of D	Generation (for Capi With Microgard™ C	llary blood Co	
	Order of Draw / Catalogue #	Additive	Recommended Fill Volumes (Min - Max)	Mix by Inverting
	Microtaixer L) 365974	K <sub>2</sub> EDTA	250µl -500µl	10x
8° (43)	MCROTANER 365965	Lithium Heparin	400µl -600µl	10x
i	365985 365985 365987	Lithium Heparin and Gel for Plasma Separation	400µl -600µl	10x
YB	MICROTAINER Bard Texes 365992	NaFI/Na <sub>2</sub> EDTA	250µl -500µl	10x
	365967 Michoranize 365978	Clot Activator and Gel for Serum Separation	200µl -400µl	5x
	365963	No Additive	400µl -600µl	Оx
Diagnostics analytical Solutions				

Picture 5

LOCAL OPERATING PROCEDURE

3



# **NEONATAL SERVICES DIVISION**

Approved by Quality & Patient Safety Committee 17 September 2020

## HEEL PRICK FOR BLOOD SAMPLING cont'd

- 11. Seal blood containers.
- 12. Apply direct pressure to puncture site until bleeding stops.
- 13. Label collection tubes with correct infant's name label.
  - NB. Must be hand written if collecting for blood group and hold. Must also be countersigned by a second clinician.
- 14. Dispose of puncture device in sharps container.
- 15. Collect and dispose of remaining equipment.
- 16. Clean blue tray and remove gloves.
- 17. Perform hand hygiene.

### 6. DOCUMENTATION

- eMR
- Neonatal Observation Chart
- Pathology Request Form

### 7. RELATED POLICIES/PROCEDURES/CLINICAL PRACTICE LOP

- NSW Health Policy Document PD2017\_013. Infection Prevention and Control Policy.
- NSW Health Guideline GL2018\_013. Work Health and Safety Blood and Body Substances
  Occupational Exposure Prevention

### 8. RISK RATING

Low

### 9. NATIONAL STANDARD

- Standard 1 Clinical Governance
- Standard 3 Preventing and Controlling Healthcare-Associated Infections
- Standard 5 Comprehensive Care
- Standard 6 Communicating for Safety
- Standard 7 Blood Management

### **10. ABBREVIATIONS AND DEFINITIONS OF TERMS**

NCC Newborn Care Centre BS	SL Blood Sugar Level
----------------------------	----------------------

### 11. REFERENCES

- ANTT® Clinical Practice Framework, Version 3.1 Copyright 2013. The Association for Safe Aseptic Practice (ASAP). www.antt.org.
- BD Asia Pacific, 2010. Capillary Blood Collection Key Aspects of Best Practice, Sources of Preanalytical Error and Laboratory Workflow Challenges. Asia Pacific Preanalytical Notes, 13 (1), p2-4.
- BD Diagnostics 2009. Capillary Blood Collection: Best Practices. Lab Notes, 20 (1), p1-5.
- CLSI H4-A5. Procedures and Devices for the Collection of Diagnostic Blood Specimen by Skin Puncture, Approved Fifth Edition, 24 (21).

### 12. AUTHOR

Primary	20.8.13	E Siddons (RN)
Revised	17.4.18	E Siddons (RN)
	8.9.20	E Siddons (RN), NCC LOPs Committee

## **REVISION & APPROVAL HISTORY**

September 2020 Revised and Approved NCC LOPs Committee April 2018 Revised and Approved Neonatal Services LOPs Group December 2013 Original Document Approved Newborn Care Centre Quality Committee

FOR REVIEW: 2025